

## Anticonvulsant Screening Program

### Test 76 Results - In-vitro Hippocampal Slice Culture Neuroprotection Assay (NP)

ASP ID: 2      U      Screen ID: 1

Solvent Code:    DMSO                      Solvent Prep:

Test Date:        26-Aug-2009

Reference:        439:178

Summary of NP Assay: NMDA

● Test Result:    No Neuroprotection

Comments:

## TEST 76: *in vitro* HIPPOCAMPAL SLICE CULTURE NEUROPROTECTION ASSAY

Compound 1 : ADD Number: 000002

Batch:

Date Started: 26-Aug-2009

Compound 2 : ADD Number:

Batch:

Date Completed: 28-Aug-2009

References: 439: 178

Excitotoxin: NMDA

Insult Duration: 4 Hours

Solvent: DMSO

Primary Screen Results: No neuroprotection observed

### EXPERIMENT IMAGES & WELL DESCRIPTION

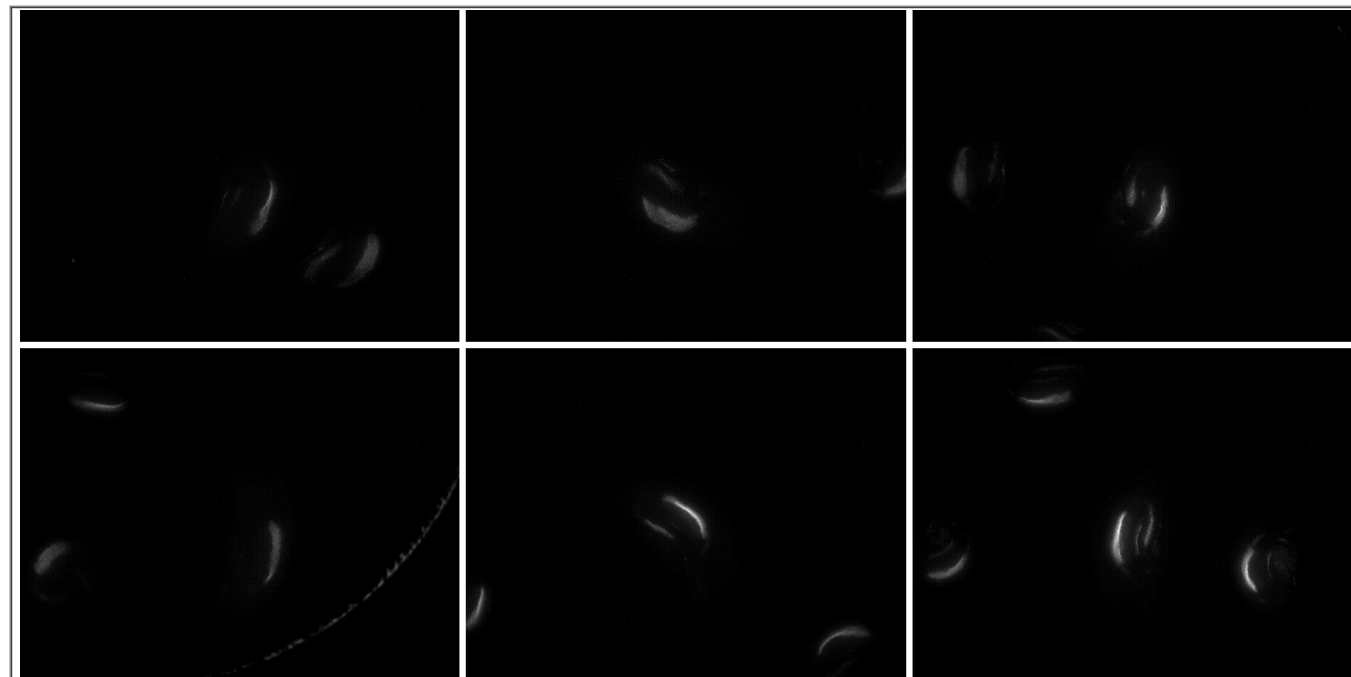
A1 NMDA 10µM

A2 NMDA 10µM +

A3 NMDA 10µM +

000002 10µM

000002 10µM



B1 NMDA 10µM

B2 NMDA 10µM +

B3 NMDA 10µM +

000002 100µM

000002 100µM

### PRIMARY SCREEN EXPERIMENT DESCRIPTION

The "Primary Screen Experiment" is a qualitative assessment of the ability of a compound to prevent excitotoxic cell death. Organotypic hippocampal slice cultures are treated with N-methyl-D-aspartate (NMDA) or kainic acid (KA) to induce neuronal cell death. Propidium iodide (PI), a membrane-impermeant compound, is included in all wells of the culture plate. Dying cells have compromised cell membranes, thus PI may diffuse into the cell, intercalate with DNA and fluoresce. Thus, the intensity of the PI fluorescence is proportional to the amount of cell death in the individual slices. Hippocampal slice cultures are treated with the excitotoxin alone, or where indicated above, with the excitotoxin and either one or two investigational compounds at the concentrations indicated. If neuroprotection occurs as a consequence of the added compound, slice cultures will have a visibly reduced fluorescent intensity when compared to the slice cultures that have been treated with the excitotoxin alone.

## Anticonvulsant Screening Program

### Test 76 Results - In-vitro Hippocampal Slice Culture Neuroprotection Assay (NP)

ASP ID: 2      U      Screen ID: 2

Solvent Code:    DMSO                      Solvent Prep:

Test Date:        26-Aug-2009

Reference:        439:178

Summary of NP Assay:    Kainic acid

● Test Result:        Neuroprotective effect

Comments:

## TEST 76: *in vitro* HIPPOCAMPAL SLICE CULTURE NEUROPROTECTION ASSAY

Compound 1 : ADD Number: 000002

Batch:

Date Started: 26-Aug-2009

Compound 2 : ADD Number:

Batch:

Date Completed: 28-Aug-2009

References: 439: 178

Excitotoxin: Kainic Acid

Insult Duration: 4 Hours

Solvent: DMSO

Primary Screen Results: Neuroprotection observed

### EXPERIMENT IMAGES & WELL DESCRIPTION

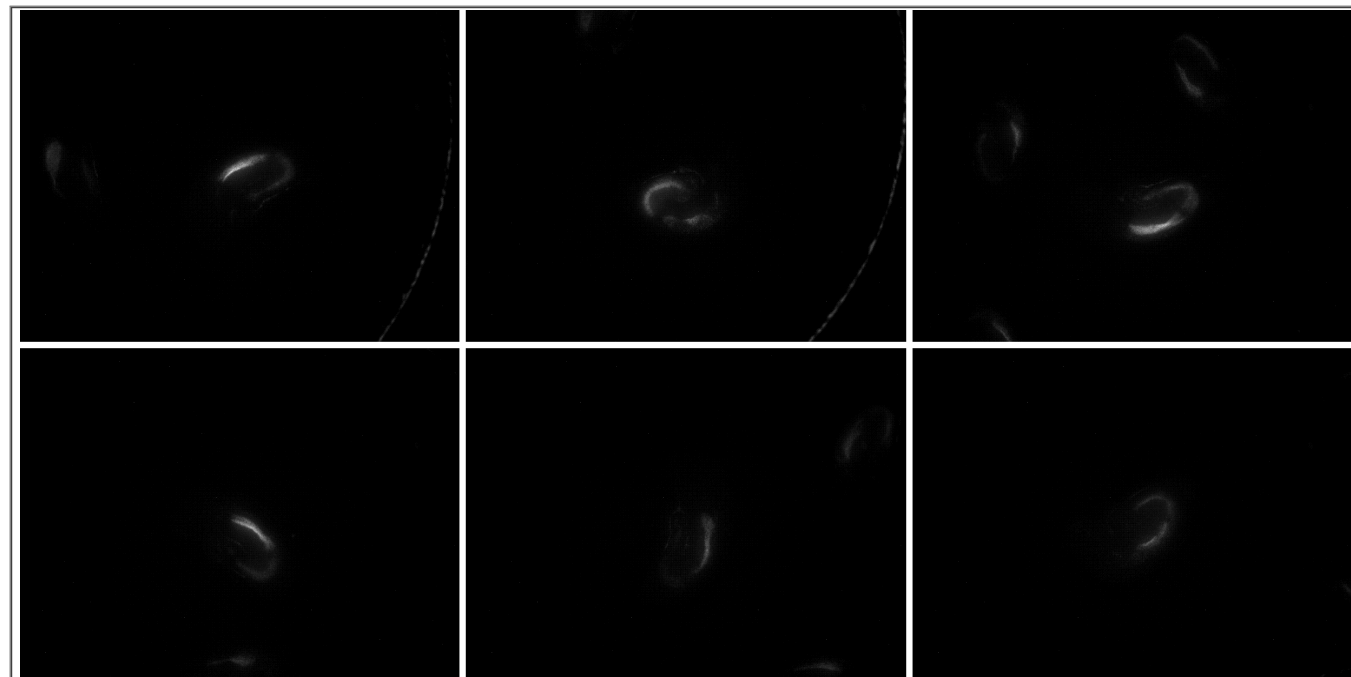
A1 KA 20µM

A2 KA 20µM +

A3 KA 20µM +

000002 10µM

000002 10µM



B1 KA 20µM

B2 KA 20µM +

B3 KA 20µM +

000002 100µM

000002 100µM

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The "Primary Screen Experiment" is a qualitative assessment of the ability of a compound to prevent excitotoxic cell death. Organotypic hippocampal slice cultures are treated with N-methyl-D-aspartate (NMDA) or kainic acid (KA) to induce neuronal cell death. Propidium iodide (PI), a membrane-impermeant compound, is included in all wells of the culture plate. Dying cells have compromised cell membranes, thus PI may diffuse into the cell, intercalate with DNA and fluoresce. Thus, the intensity of the PI fluorescence is proportional to the amount of cell death in the individual slices. Hippocampal slice cultures are treated with the excitotoxin alone, or where indicated above, with the excitotoxin and either one or two investigational compounds at the concentrations indicated. If neuroprotection occurs as a consequence of the added compound, slice cultures will have a visibly reduced fluorescent intensity when compared to the slice cultures that have been treated with the excitotoxin alone.

## Anticonvulsant Screening Program

### Test 76 Results - In-vitro Hippocampal Slice Culture Neuroprotection Assay (NP)

ASP ID: 2      XX      Screen ID: 3

Solvent Code: DMSO      Solvent Prep:

Test Date: 26-Aug-2009

Reference: 439:178,228,234

Summary of NP Assay: Kainic acid

● Test Result: No Neuroprotection

#### Response

Excitotoxin: Kainic acid

| ASP Compound Conc.(uM) | # of slices | % of Total Propidium Iodide Uptake<br>(mean +/- S.E.M) |                          |
|------------------------|-------------|--------------------------------------------------------|--------------------------|
| 0                      | 8           | 52.1 +/- 4.1                                           | <input type="checkbox"/> |
| 10                     | 8           | 49.9 +/- 4.4                                           | <input type="checkbox"/> |
| 100                    | 7           | 46.8 +/- 5.5                                           | <input type="checkbox"/> |

Note: box is checked if data is significantly different from excitotoxin treatment alone, p<0.05.

IC50(uM): > 100.00 +/- S.E. M

Comments:

## TEST 76: *in vitro* HIPPOCAMPAL SLICE CULTURE NEUROPROTECTION ASSAY

Compound 1 : ADD Number: 000002

Batch:

Date Started: 26-Aug-2009

Compound 2 : ADD Number:

Batch:

Date Completed: 30-Oct-2009

References: 439: 178, 228, 234

Excitotoxin: Kainic Acid

Insult Duration: 4 Hours

Solvent: DMSO

Primary Screen Results: No neuroprotection observed

### EXPERIMENT IMAGES & WELL DESCRIPTION

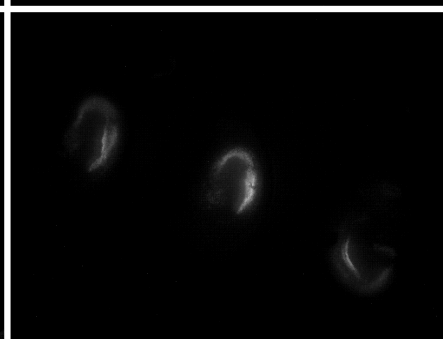
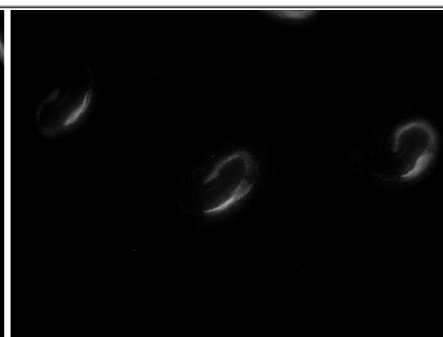
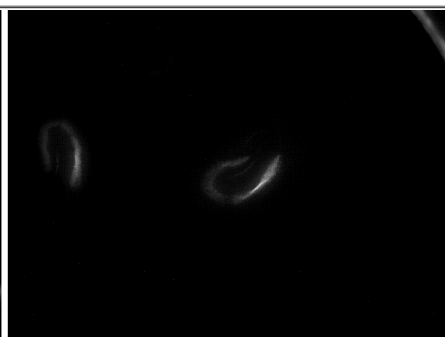
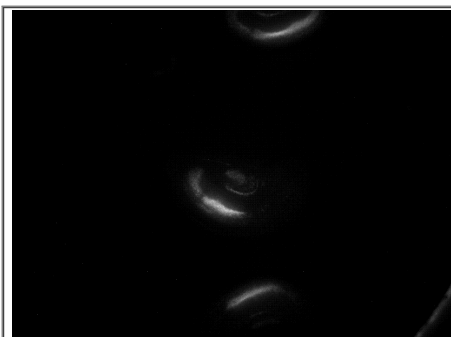
A1 KA 20µM

A2 KA 20µM +

A3 KA 20µM +

000002 10µM

000002 10µM



B1 KA 20µM

B2 KA 20µM +

B3 KA 20µM +

000002 100µM

000002 100µM

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The "Primary Screen Experiment" is a qualitative assessment of the ability of a compound to prevent excitotoxic cell death. Organotypic hippocampal slice cultures are treated with N-methyl-D-aspartate (NMDA) or kainic acid (KA) to induce neuronal cell death. Propidium iodide (PI), a membrane-impermeant compound, is included in all wells of the culture plate. Dying cells have compromised cell membranes, thus PI may diffuse into the cell, intercalate with DNA and fluoresce. Thus, the intensity of the PI fluorescence is proportional to the amount of cell death in the individual slices. Hippocampal slice cultures are treated with the excitotoxin alone, or where indicated above, with the excitotoxin and either one or two investigational compounds at the concentrations indicated. If neuroprotection occurs as a consequence of the added compound, slice cultures will have a visibly reduced fluorescent intensity when compared to the slice cultures that have been treated with the excitotoxin alone.